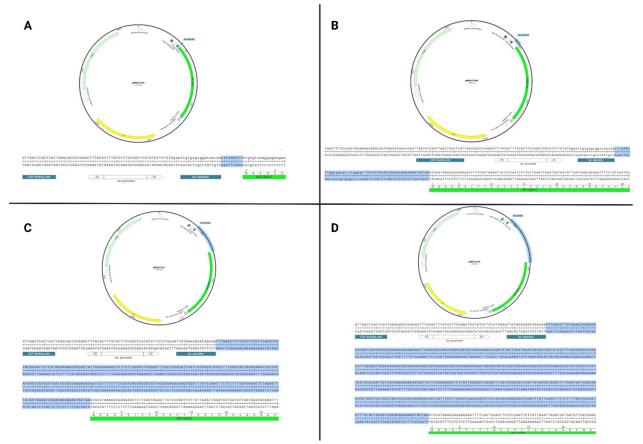
## SUPPLEMENTAL MATERIAL

**TABLE. S1 Inverse PCR primers for amplification of pEAH23A.** The sequences and melting temperature of all the primers used for inverse PCR as designed using SnapGene and NEBase changer.

Primer Name	Sequence (5'>3')	Tm (°C)
pBRAT_rev	TAAATTGTTATCCGCTCA CAATTC	60°C
pBRAT_fwd_a	AGCTTATGCGTAAAGGA GAAGAAC	60°C
pBRAT_fwd_b	AGCTTGGTGGCACTCTA GAG	58°C
pBRAT_fwd_c	AGCTTTTGTCTTCCTTGA CGTC	58°C
pBRAT_fwd_d	AGCTTGTAGACCACGGC GAA	58°C

**TABLE. S2 pEAH23A amplified regions and respective primers.** The primer pairs that were utilized to amplify the backbone of pEAH23A to generate each amplicon.

Amplicon Name	Primers	Size (bp)
pBRAT24A	pBRAT_rev, pBRAT_fwd_a	3153
pBRAT24B	pBRAT_rev, pBRAT_fwd_b	3199
pBRAT24C	pBRAT_rev, pBRAT_fwd_c	3420
pBRAT24D	pBRAT_rev, pBRAT_fwd_d	3643



**FIG. S1 SnapGene analysis shows varying sequences of interest in the pBRAT24 plasmids.** Plasmid maps and the sequences between the *lac* operator and *gfp* for each of the pBRAT24 constructs: (A) pBRAT24A, (B) pBRAT24B, (C) pBRAT24C, (D) pBRAT24D.

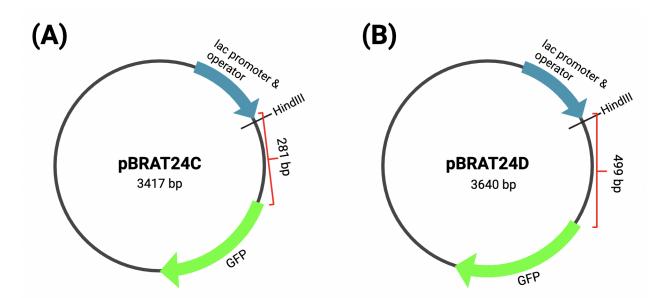
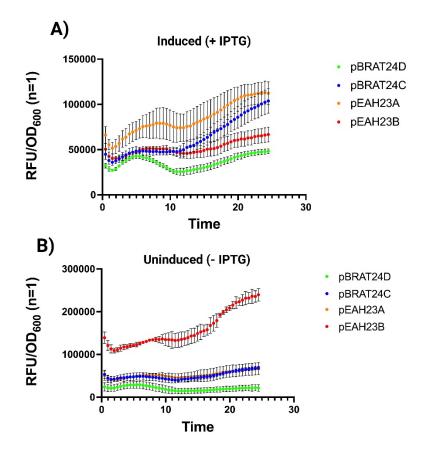


FIG. S2 Plasmidsaurus sequencing shows successful construction of pBRAT23C and pBRAT24D. Diagrams depicting a simplified representation of the Plasmidsaurus sequencing results of (A) pBRAT24C diluted to 30 ng/ $\mu$ L and (B) pBRAT24D diluted to 30 ng/ $\mu$ L.



**FIG. S3 Intermediate deletions of pEAH23A led to no increase in RFU/OD600 over time.** Results of **(A)** IPTG or **(B)** distilled water induction of overnight liquid cultures as measured by a fluorescent plate reader (*BioTek*). Colonies from pEAH23A, pEAH23B, pBRAT24C, pBRAT24D, and pDO6935 were initially grown overnight at 37°C. All overnight cultures were then seeded into a 96 well plate with either IPTG or distilled water in triplicate and grown again at 37 °C. GFP measurements were normalized to OD600 and standardized to the negative control: pDO6935. Each dot represents a time point in which fluorescence was measured. Results were plotted in GraphPad Prism<sup>TM</sup>. Error bars show standard error mean of triplicate measurements (n=1).