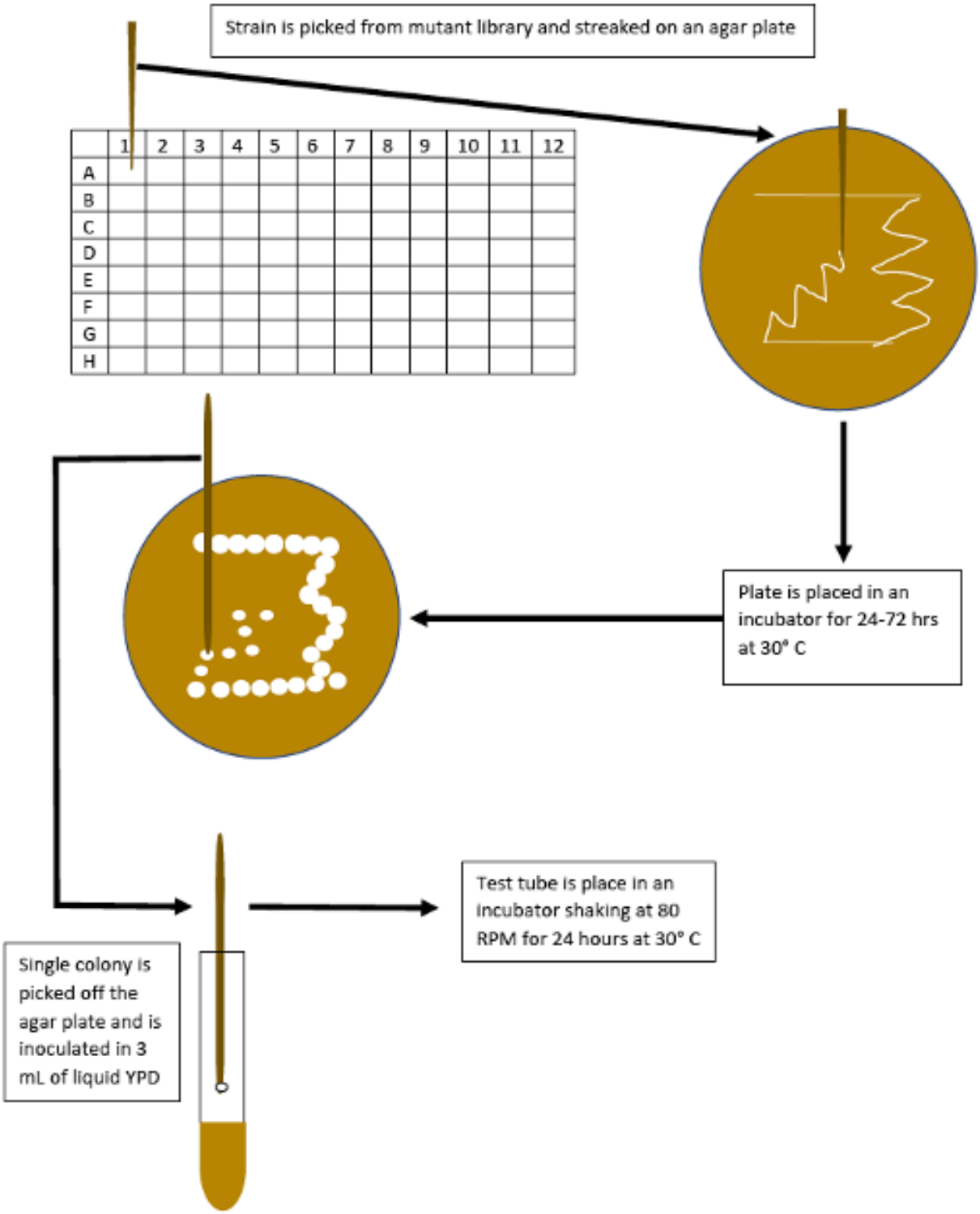
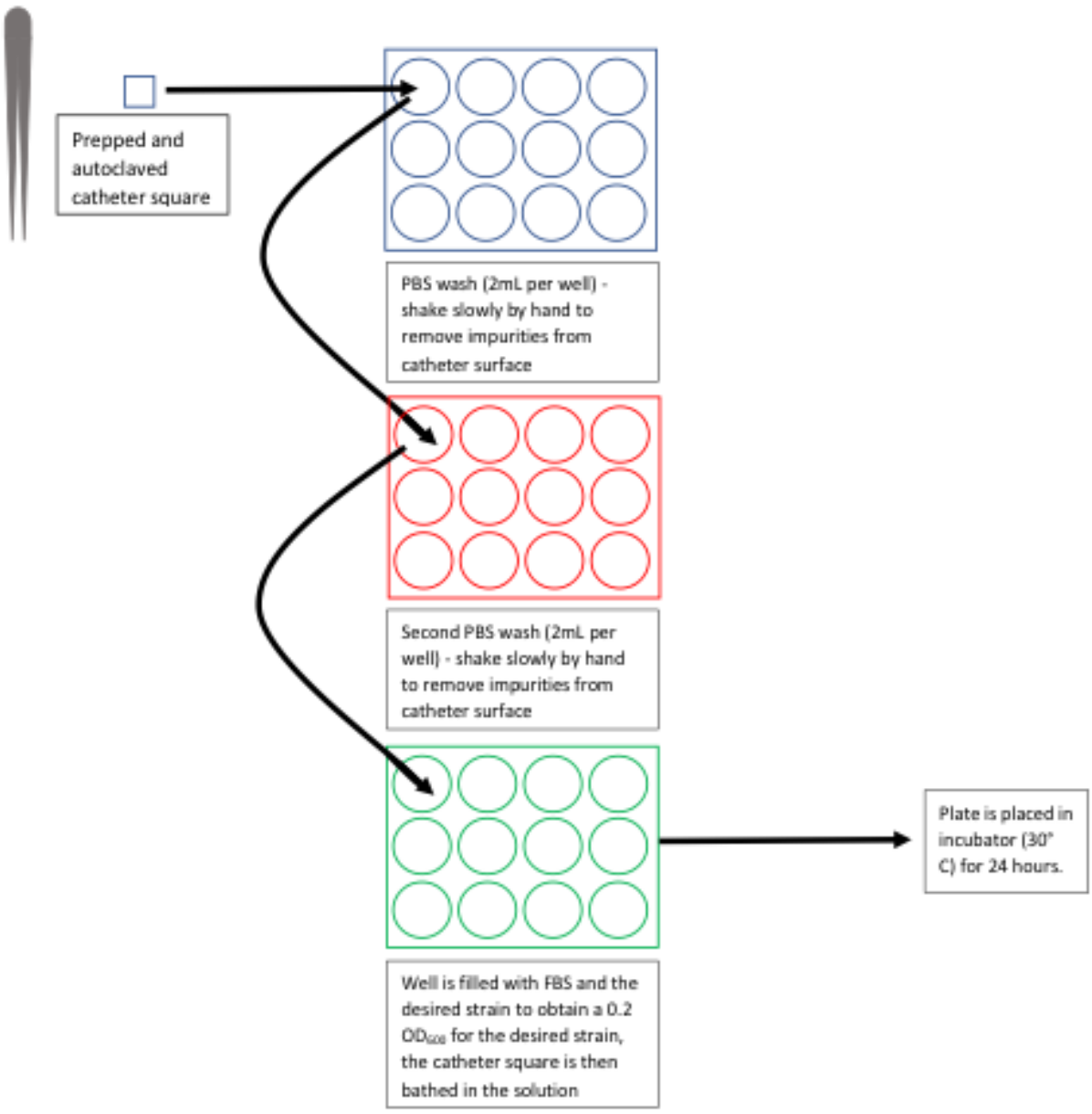
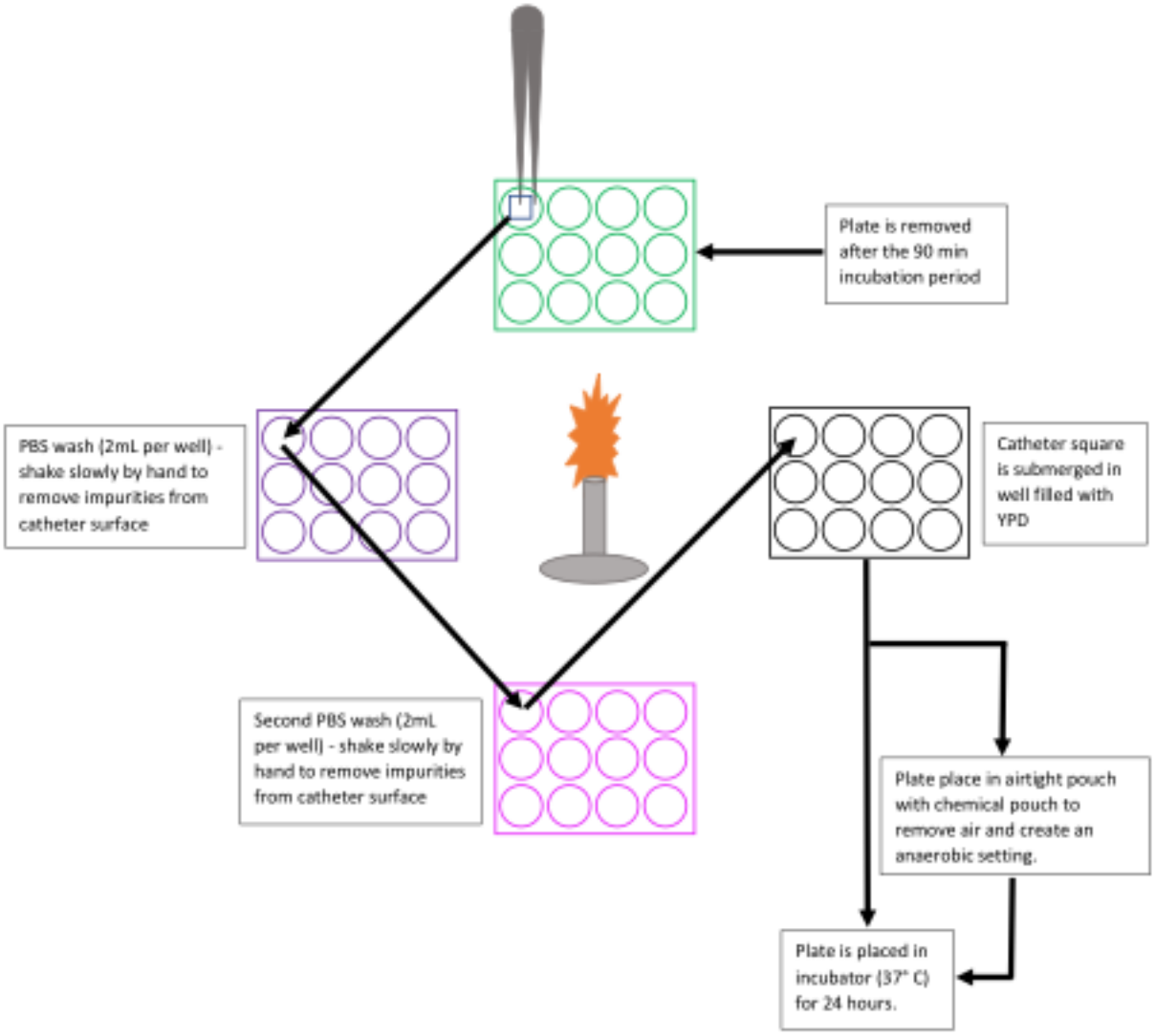


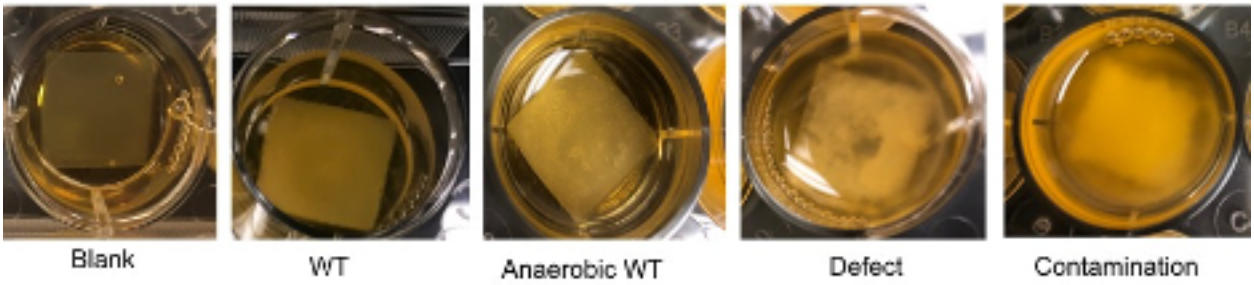
Supplemental Figure 1 Setting up cultures. Schematic overview of how to remove culture from the freezer, streak for single colonies, pick a single colony, and add to the liquid medium.



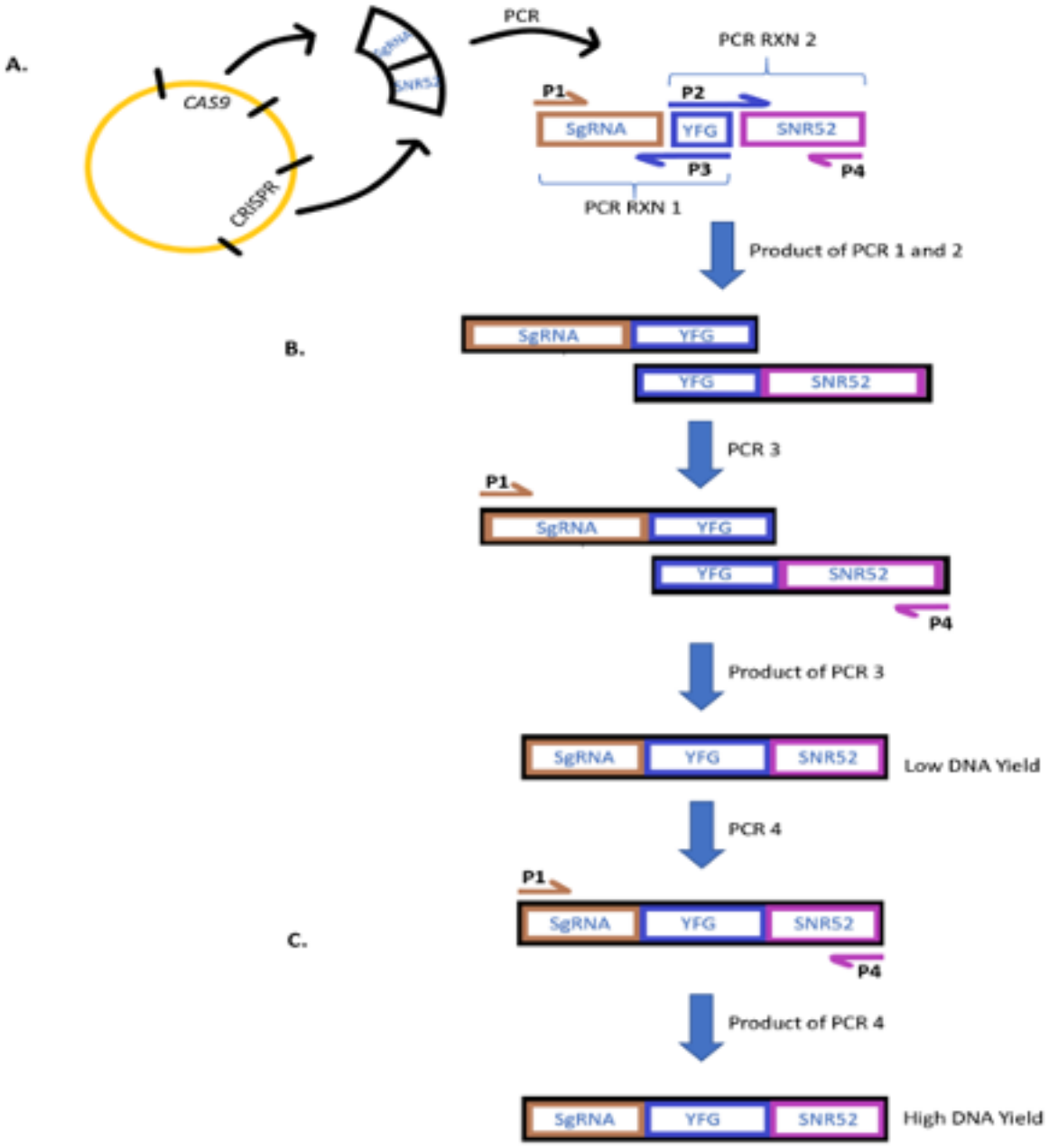
Supplemental Figure 2. Schematic overview and order for how to transfer and wash catheter squares for preparation for incubation with FBS.



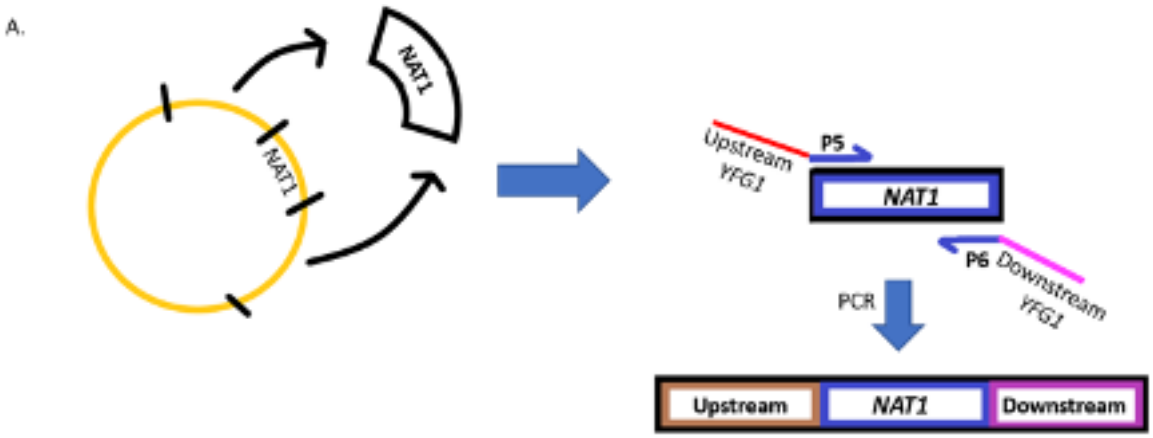
Supplemental Figure 3 Schematic overview and ordering for transferring and washing catheter squares for preparation for incubation. Also a map of what to do.



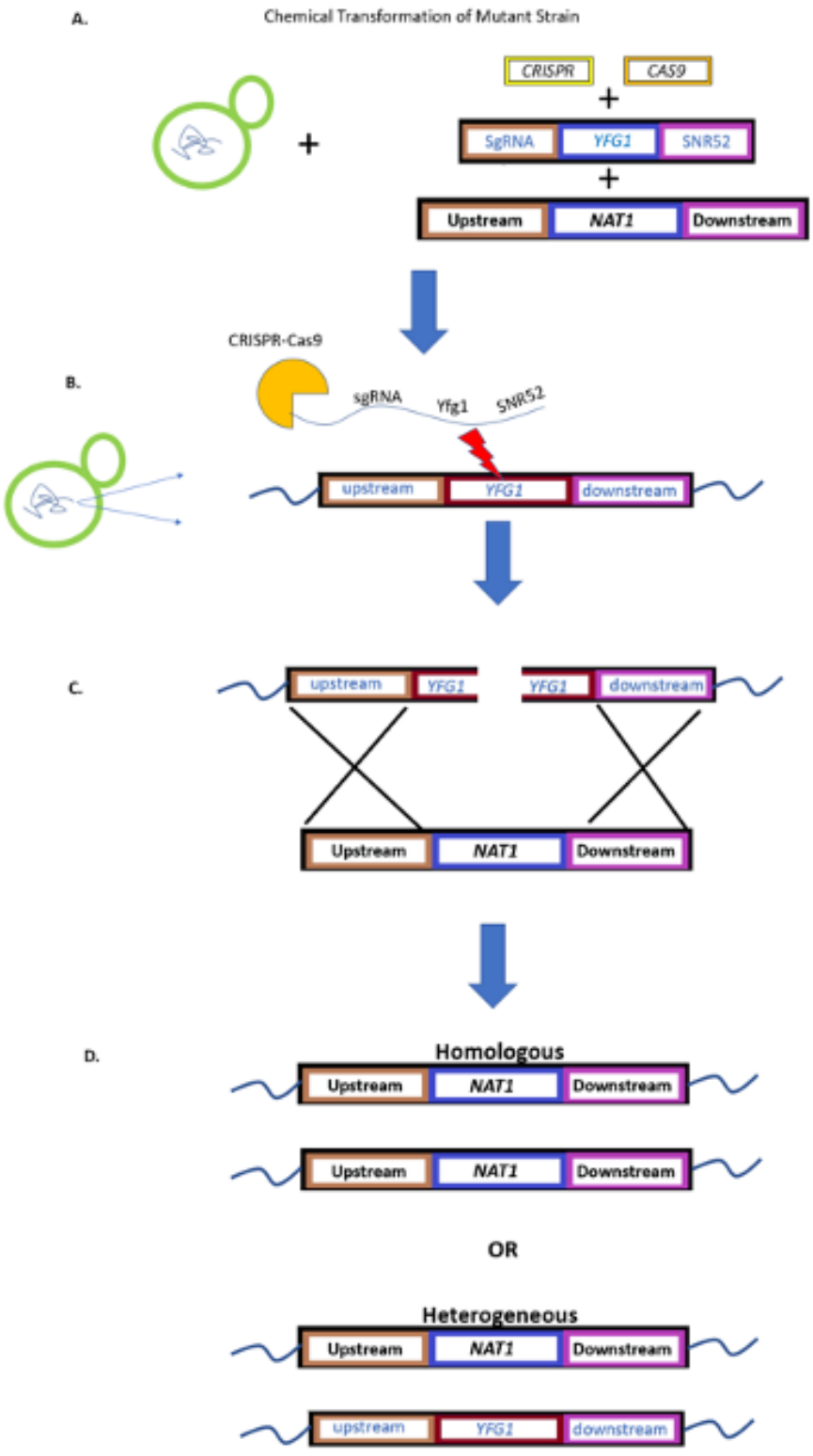
Supplemental Figure 4 Examples of what the wells can look like in different conditions. The first well shows the appearance of the negative culture well. The second well shows the appearance of anaerobic wild type biofilm. The third shows the appearance of an anaerobic wild type biofilm. Well 4 shows the appearance of a typical mutant for biofilm formation. Well 6 shows the appearance of what many contamination wells look like.



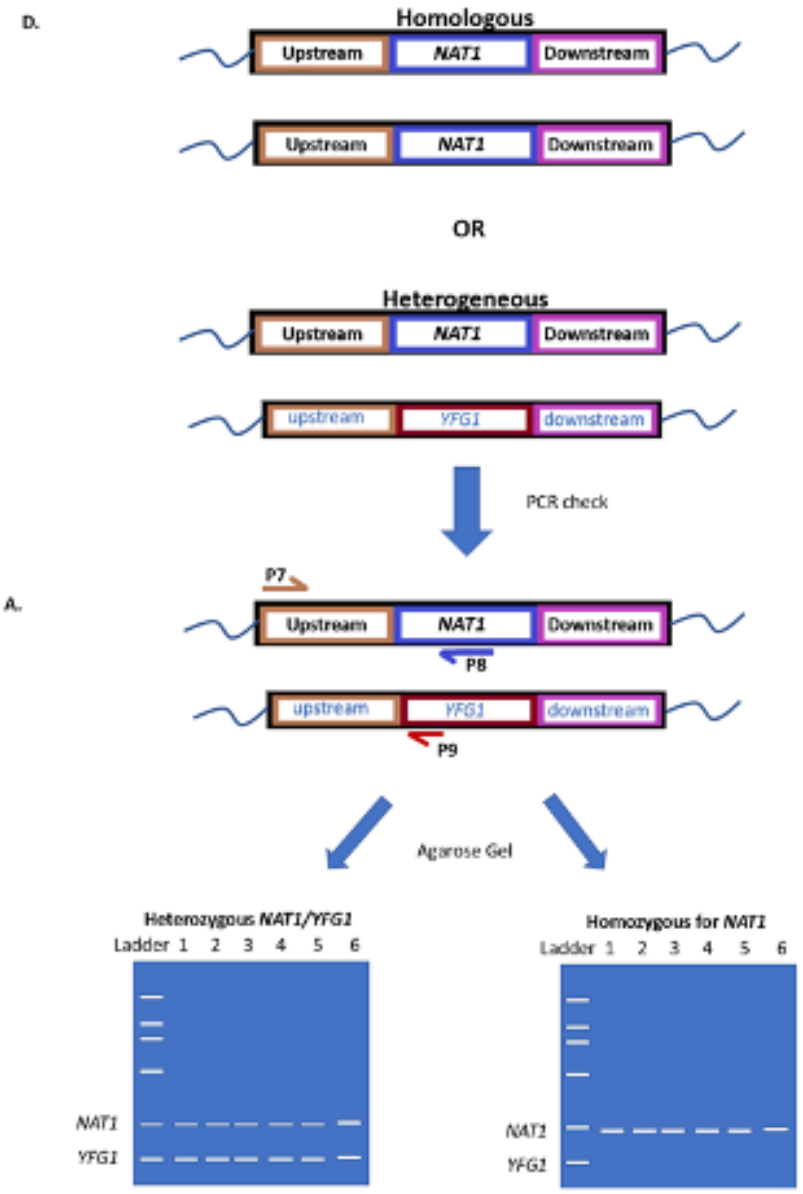
Supplemental Figure 5: Schematic overview and ordering for the process of transforming the guide sequence (*snr52, SgRNA*, and *YFG1*) via PCR.



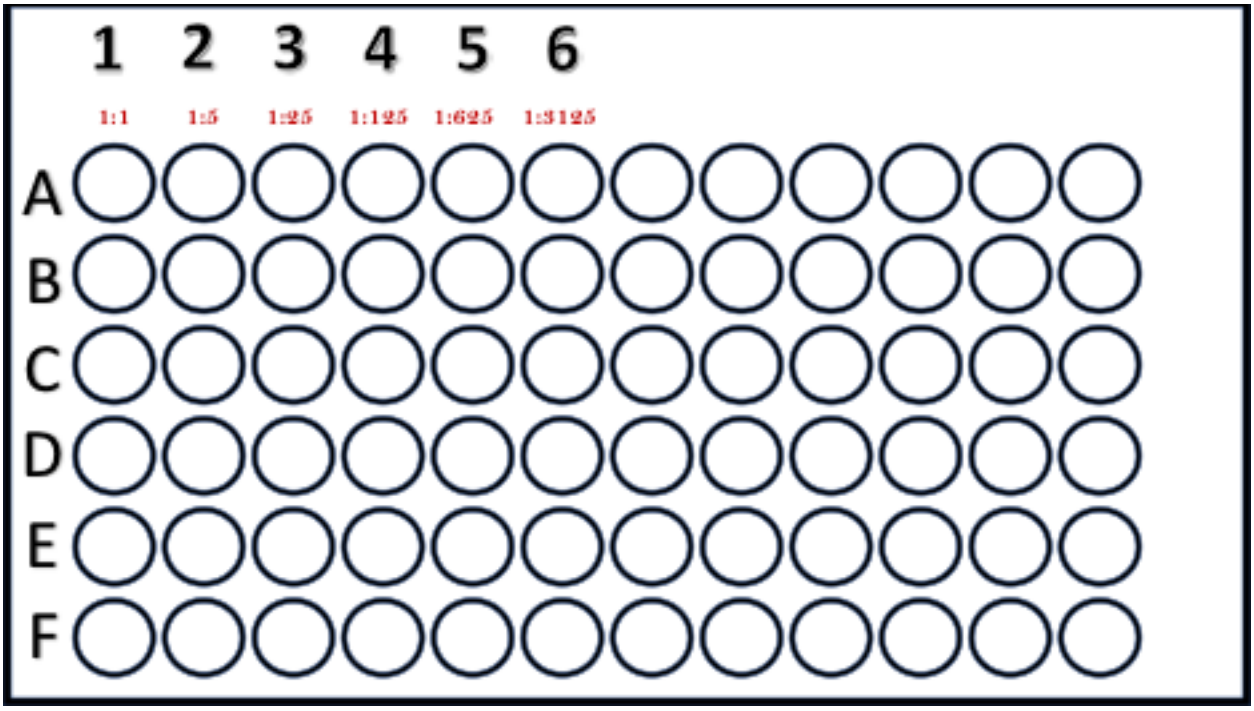
Supplemental Figure 6: Schematic overview and order for the process of creating the *NAT1* construct. This construct will be used to aid in the deletion process. This construct will allow for an assay on nourseothricin plates, which will reveal if *YFG1* has been removed or is still present.



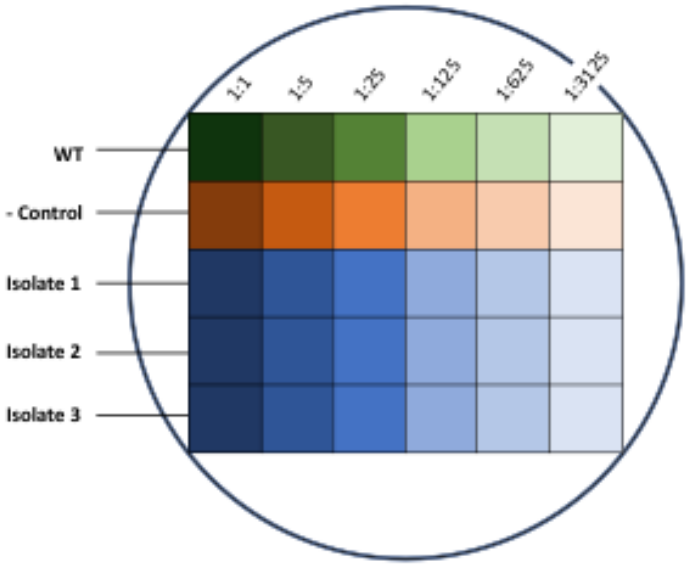
Supplemental Figure 7: Schematic overview and ordering for the process of transforming the CRISPR Cas9 DNA with SgRNA- *YFG1* -SNR52 and *NAT1*.



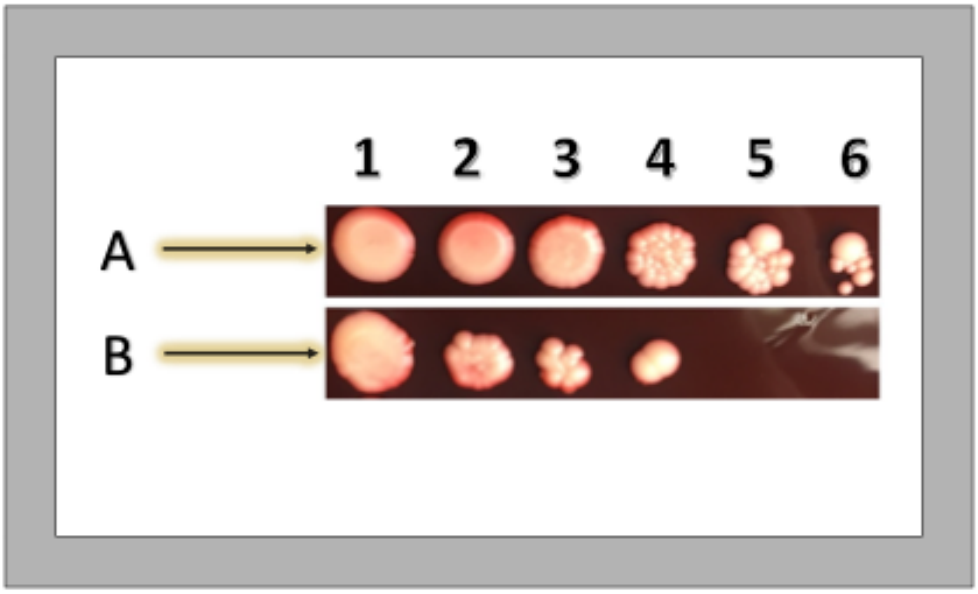
Supplemental Figure 8: Schematic overview and order for how to use PCR to check colonies to determine the heterogeneous and homogenous products using an agarose gel.



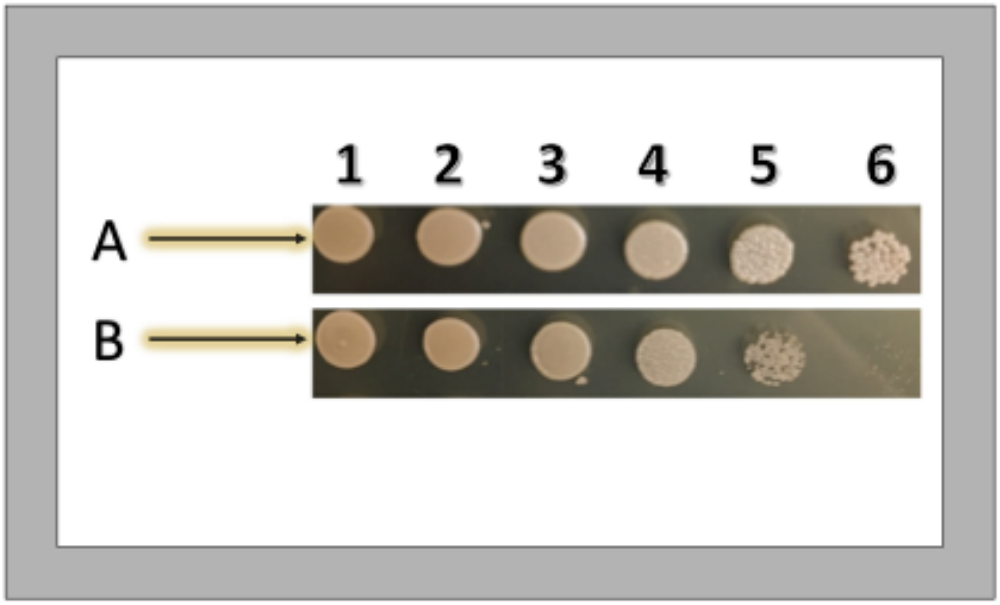
Supplemental Figure 9: 96 Well Plate Sample. In red includes proper dilutions after transfer between lanes.



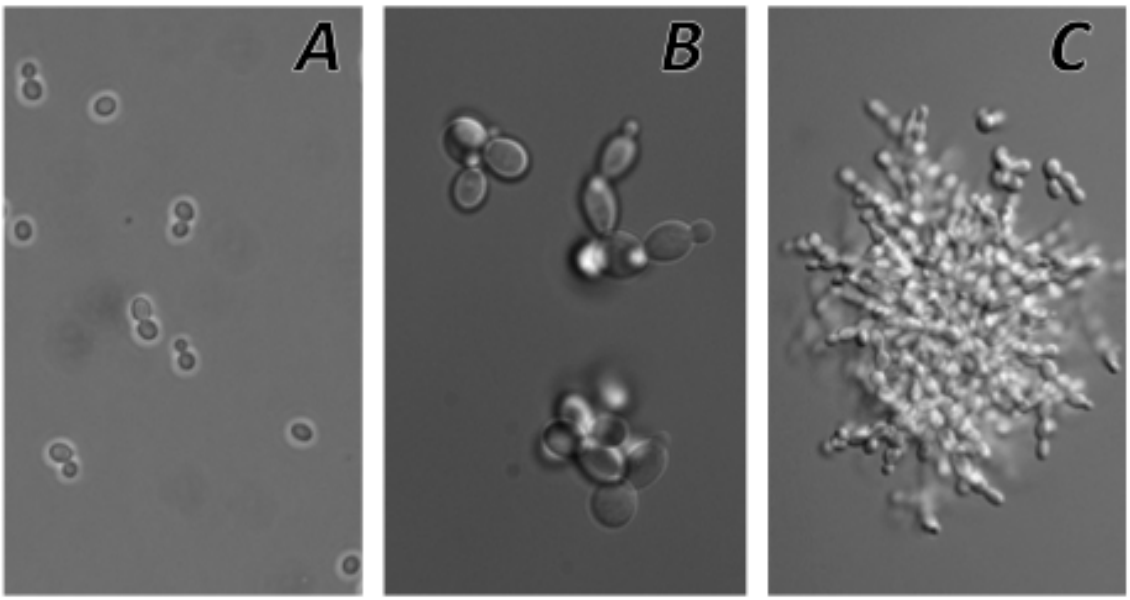
Supplemental Figure 10: Spot Plate Sample. Each sample has been transferred from the 96 well plate in the decreasing dilution (rows). Each row is also specific to the strain type; WT, - control, isolate 1, isolate 2, and isolate 3.



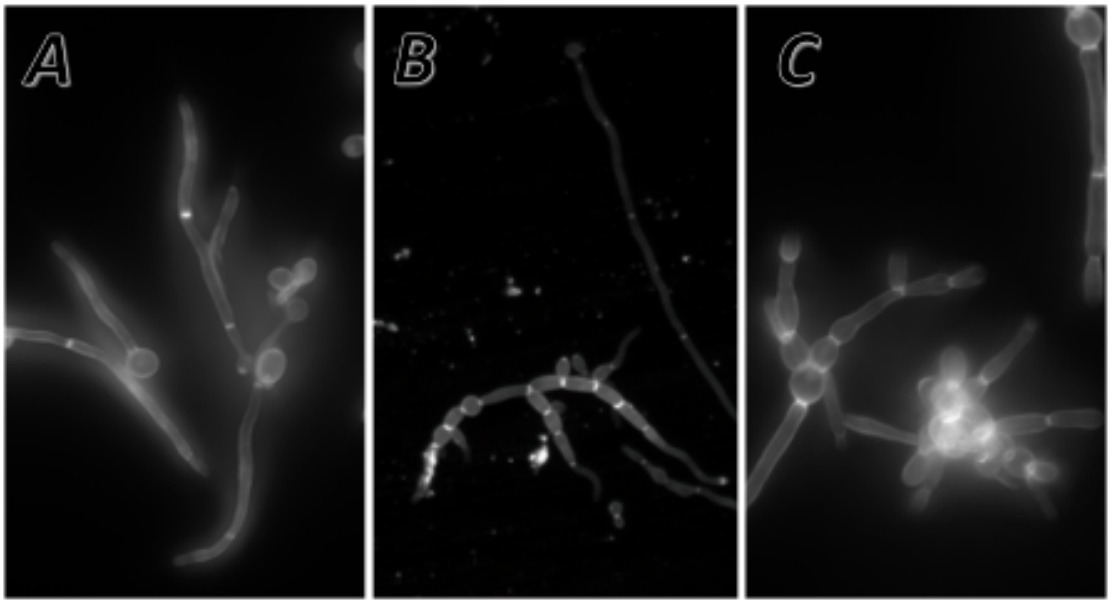
Supplemental Figure 11. Spot Plate Sectional Congo Red- A) Wild-type strain growing successfully at all dilutions. B) Example of isolate strain high density of culture in lane 1 dramatically decreases and disappears in lane 5 & 6 (Isolate unpublished mutants).



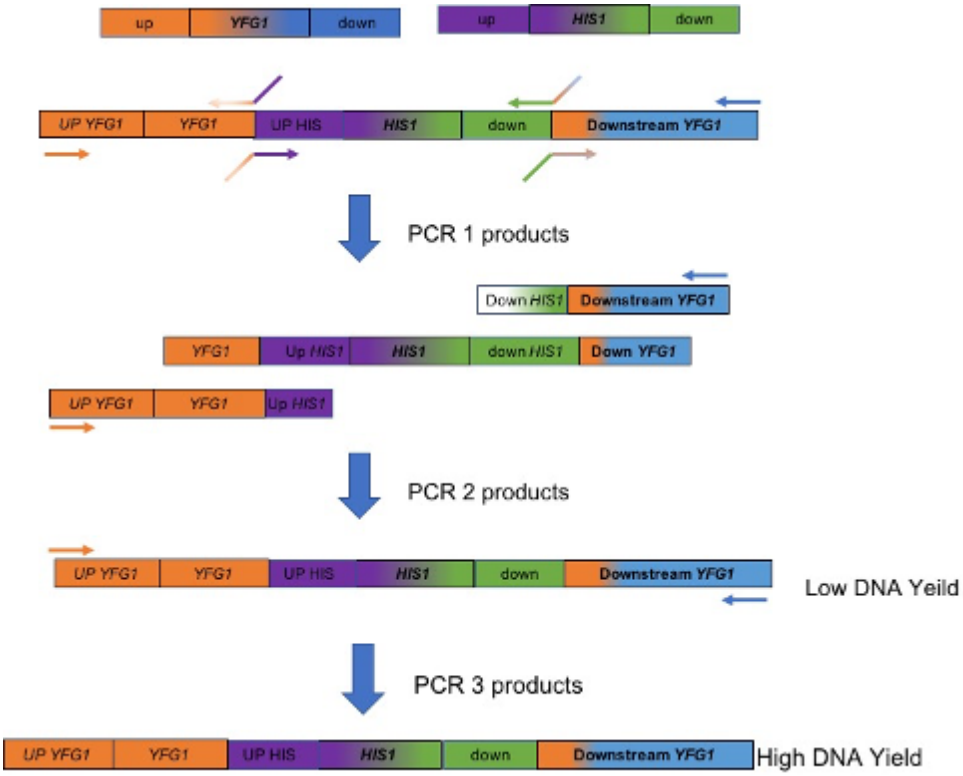
Supplemental Figure 12: Spot Plate Sectional NaCl Assay- A) Wild-type DAY286 growing on NaCl plate density diminishes, but the formation is still prevalent in lane 5. B) Example of isolated growth, lanes dramatically reduce between lanes 4-6 (Isolate unpublished mutants).



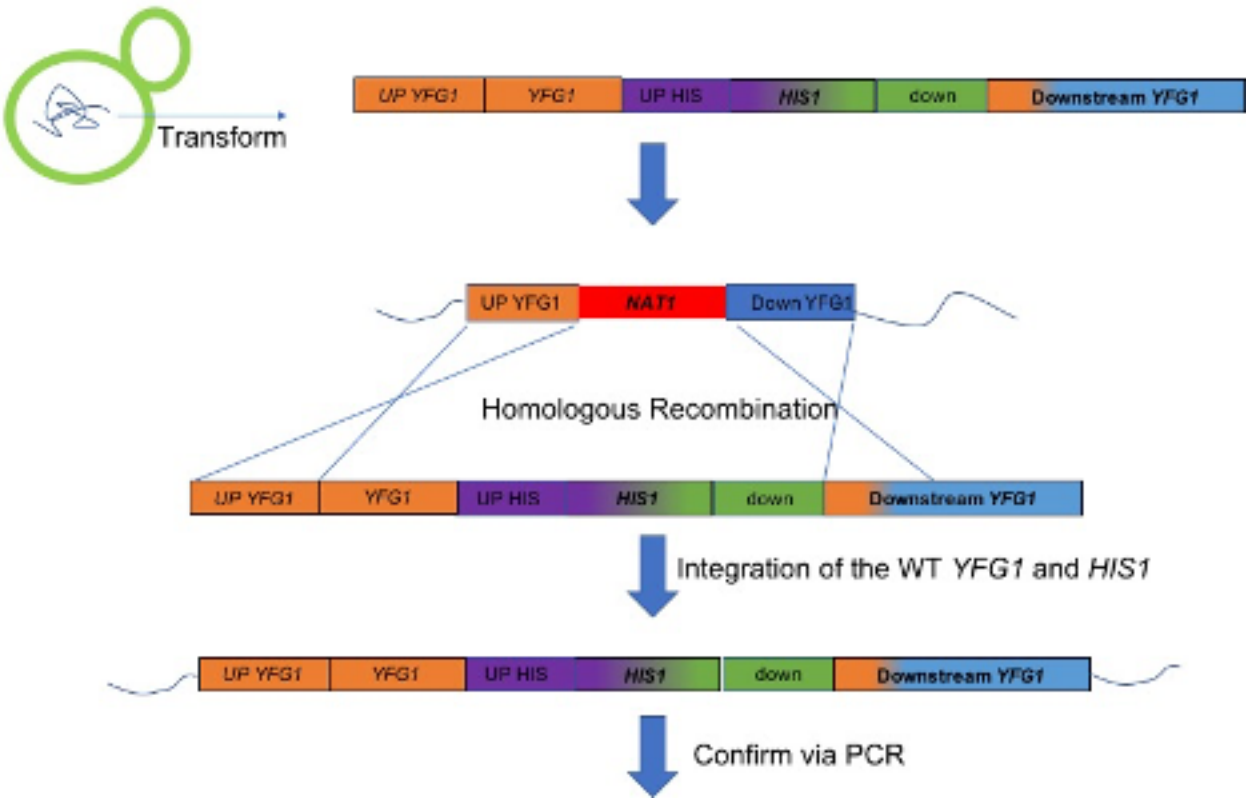
Supplemental Figure 13: Yeast Formation Test - A) Wild-type DAY286 yeast budding B&C) Abnormal yeast budding seen in an *SNF5* deletion strain.



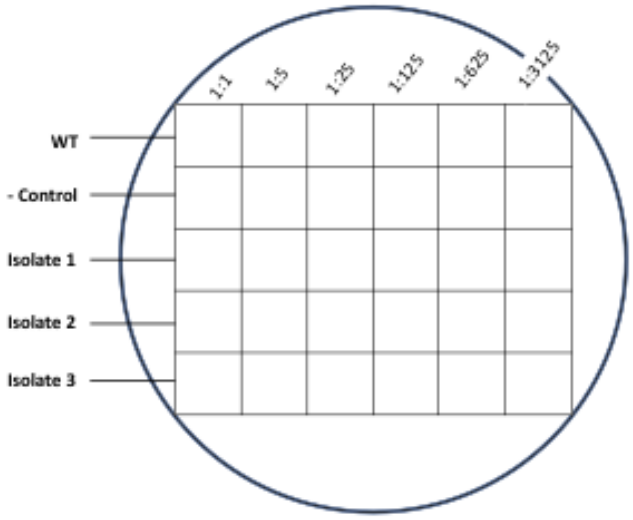
Supplemental Figure 14: Hyphae Formation- A) Wild-type DAY286 hyphae formation B&C) Possible types of abnormal hyphae formation (unpublished mutant data).



Supplemental Figure 15 Schematic overview and ordering for creation of complement construct.



Supplemental Figure 16 Schematic overview and order for what happens during transformation and how to confirm results.



Supplemental Figure 17: Printable stencil to assist with spot plate technique.